Meerut Institute of Technology and Engineering

DST – Fist Center
Central Facility – Flow Cytometer

Requisition form for using Flow Cytometer facility

instrument.dbt@miet.ac.in

Date: .....

1) Name of the User*:

2) Name of the Organization*: .....

3) Name of the HOD/ Principal Investigator*: .....

4) User's Email ID and Tel No*: .....

5) Experiment Name:

6) Experiment type: Acquisition and data analysis

7) Name of Control:

8) Name of Sample:

9) Size of Cells/Particles:

10) Number of parameters to be analysied:

11) Name of the fluorophores:

12) Preferred date of the slot for experiment:

Optical Filter and Acquisition Selection for the Experiment

3 Blue 1 Red Configuration (Standard Filters)			3 Blue 1 Red Configuration 2			2 Blue 2 Red Configuration		
Detector	**Filter**	**Fluorochrome**	**Detect**	**Filter**	**Fluorochrome**	**Detector**	**Filter**	**Fluorochrome**
FL1	530/30	FITC, GFP, CFSE	**FL1**	530/30	FITC, GFP, CFSE	**FL1**	530/30	FITC, GFP, CFSE
FL2	585/40	PE, PI, PE-Texas Red	**FL2**	585/40	PE	**FL2**	585/40	PE, PI, PE-Texas Red
FL3	610/20	PerCP-Cy5.5 PE-Cy5, PE-Cy7	**FL3**	610/20	PI, PE-Texas Red	**FL3**	780/60	APC-Cy™7 (and equivalents)
FL4	675/25	APC, Alexa-647, PE-Cy5	**FL4**	675/25	APC, Alexa-647, PE-Cy5	**FL4**	675/25	APC (and equivalents)

NOTE: When operating in the *3 blue 1 red* configuration, place 610/20 eitherbandpass or the 670 LP filtersin position FL3. For best results when analyzing PE and PE-Texas Red (PE-TR) simultaneously, select thefilters with the following signal-intensity considerations in mind:

- PE-bright, PE-TR-moderate to bright: FL3 = 670 LP
- PE-dim to moderate, PE-TR any level: FL3 = 610/20
- PE-bright, PE-TR dim: may be difficult to separate; consider using the *four blue* configuration with a630/30 in FL4 to detect PE-TR.

Other panel selection is also possible depending your experiment requirements.

If not sure about selection of filters & laser then list down the Excitation & Emission spectra of the dye / fluorophore.

Additional information: (Constraints/Preferences/ etc.)

— I understand that the samples will be analyzed according to the choices recorded in the form.

- KINDLY ENSURE THAT THE GIVEN SAMPLE IS NOT INFECTIOUS OR POISONOUS OR TOXIC IN ANY WAY
- Whenever the results are used in the publications, appropriate acknowledgment of usage of MIET Meerut's Flow Cytometer facility must be mentioned. The details can be forwarded to instrument.dbt@miet.ac.in

Important information:

1) Acquisition refers to analyzing samples on Flow Cytometer' refers to analyzing desired population.
0) Gating of population is experiment design specific and should be suggested by the user.
1) Gating and data analysis can also be done offline by the user.
2) Instrument is equipped to normally handle cells/particles in the size range 0.2 to 25 microns. While instrument cannot "strictly" handle cells/particles above 25 microns, it may be possible to analyze samples of size less than 0.2 microns. Such samples, that is, those containing particles less than 0.2 microns will be handled on a case-by-case basis and user requesting analysis of such samples must discuss with the operator at flowcytometer@miet.ac.in prior to submitting a request. Duly filled form shall be sent to flowcytometer@miet.ac.in.
3) MIET Meerut shall not be responsible for the loss of sample due to breakdown or any other technical difficulties.
4) In case of breakdown or technical difficulties, if the samples are not analyzed on the intended date, the slot shall be rescheduled and the user will be informed via email about the date and time of the slot.
5) Attachments like reference papers; technical data sheet etc. can be sent in mail to provide any additional information about the experiment.

I have read and understood above me[illegible]